







used. Superior detection of miR-16 was obtained using Norgen's modified protocol with a smaller standard deviation. Moreover, Ct values obtained using Norgen's protocol were correlated with the amount of input (i.e. higher input = lower Ct value) indicating the RNA was of high purity and lacked PCR inhibitors. In contrast, the phenol-based method had a higher average Ct value, and did not correlate with input volume, indicating possible PCR inhibition.

**Figure 6.** Comparison of Relative PCR Inhibition Observed Between Norgen's Total RNA Kit (With Modifications) and the Competitor's Phenol-Based Method for Plasma microRNA Isolation. Total RNA including microRNA was isolated from 200 µL of sheep plasma collected in EDTA tubes and spiked with human microRNAs. Equal portions (4, 8 or 12 µL) of isolated RNA were then used for detection of miR-16 using RT-qPCR.

## CONCLUSIONS

From the data presented in this report, the following can be concluded:

1. Norgen's silicon carbide technology provides an efficient and effective way of isolating plasma miRNA without the use of hazardous organic extraction. The resulting advantages includes:
  - a. **Better Consistency**
  - b. **Better Recovery**
  - c. **Faster Protocol**
  - d. **High-Throughput (such as 96-wells)**
2. Norgen's silicon carbide technology provides a solution to overcome the main problem associated with plasma miRNA purification - sample source. A consistent microRNA isolation procedure, without the use of phenol is provided for plasma samples collected

into various anticoagulants such as Citrate, EDTA and Heparin.

3. **No PCR inhibition** was observed with Norgen's silicon carbide-based plasma microRNA protocol, even for samples purified from blood tubes containing heparin, a known PCR inhibitor.

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